

In Vitro Elution Characteristics of Gentamicin and Vancomycin from Synthetic Bone Graft Substitutes

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Abstract: *Objects:* Beta tricalciumphosphate pellets loaded with individualized antibiotics may represent novel options in the treatment of osteomyelitis and infectious bone disease. Here, the *in vitro* antibiotic elution of vancomycin and gentamicin from the synthetic bone graft substitutes Cerasorb[®] and Cerasorb M[®] was tested.

Methods: Antibiotic elution and concentration of gentamicin and vancomycin were measured using photometrically-based measurement and homogeneous particle-enhanced turbidimetric inhibition immunoassays (PETINIA).

Results: Initially both materials showed a high release of the loaded antibiotics, with Cerasorb M[®] showing lower release levels for gentamicin and vancomycin than Cerasorb[®]. Gentamicin concentrations of Cerasorb M granules and Cerasorb were below the minimum detection threshold until day four and six of the experiment respectively. The vancomycin release-level followed a similar pattern, although the vancomycin concentration eluted by Cerasorb M[®] granules stayed above the detection threshold during the experimental time.

Conclusions: Cerasorb[®] and Cerasorb M[®] may represent a new treatment option in osteomyelitis and infectious bone disease.

Keywords: Cerasorb[®], Cerasorb M[®], Osteomyelitis, beta tricalciumphosphate.

1. INTRODUCTION

The treatment of osteomyelitis remains a challenging problem for orthopaedic surgeons. Reaching a chronic state of osteomyelitis, the local vascularity is compromised. Therefore obtaining an effective local antibiotic concentration is hardly gained through oral or parental administration. Based on this knowledge, the local application of antibiotics, besides surgical debridement, represents a viable alternative treatment to handle osteomyelitis by reaching higher local antibiotic concentrations and minimizing systemic side effects [1].

Since the introduction of antibiotic-impregnated composite and chains, the local application of antibiotics is a common and widely used procedure in the fields of trauma and orthopaedic surgery [2, 3].

The main disadvantages in the utilisation of antibiotic-impregnated chains are the necessity of a second surgery to remove the material, as well as not being able to provide individualized chemotherapy in terms of local administration of different antibiotics.

Today there are several bone-graft substitutes available, which can be used as a controlled release delivery system for antibiotics. E.g. the usage of calcium sulfate as a bone graft substitute for bone lesions is known for a long time, with proven efficiency in experimental and clinical trials [4-7]. Adversely several trials showed a transient cytotoxic effect of calcium sulphate, resulting in inflammatory reactions [8-10].

Nevertheless in several trials calcium sulphate was used as an antibiotic-carrier material and has proven its efficiency as well as its security as a carrier substance [11, 12].

The major drawback in earlier studies of calcium sulphate as an antibiotic-carrier material was the addition of the appropriate antibiotic to the calcium sulphate before

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hardening, creating possible risks of impairing the antibiotic activity due to hardening or sterilization procedures [11].

Today there is a formulation of the material solving this problem by loading the antibiotic after sterilization, directly before application [13].

Due to the cytotoxic effect of pure calcium sulphate and the probable loss of antibiotic efficiency during hardening, several combinations of calcium sulphate with different materials were developed, e.g. the combination of calcium sulphate and nanocrystalline hydroxyapatite. This combination is available in different moulds, which can be loaded with antibiotics after sterilization or hardening. Over a period of 10 days the composite material of calcium sulphate and hydroxyapatite showed a nearly complete release of the loaded antibiotics. Within these 10 days of trial the composite material revealed high initial antibiotic release with subsequent decline. The different release of antibiotics between the combination of a calcium sulphate/hydroxyapatite mixture and pure calcium sulphate was compared in several studies. Such studies found a higher early release (during the first 5 days) for the calcium sulphate/hydroxyapatite mixture. After 5 days of trial pure calcium sulphate showed a higher antibiotic release [14].

The same studies also showed an excellent resorption rate, biocompatibility and antibiotic release for the calcium sulphate/hydroxyapatite composite material.

Several other materials were shown to display similarly high levels of biocompatibility, resorption rate and osseointegration. Widely available are calcium phosphates, mainly hydroxyapatite and tricalcium phosphate. Both materials are reasonable alternatives for the current gold standard in the treatment of bone lesions, the autogenic cancellous bone repair [15, 16].

The aim of this study was to evaluate the applicability of Cerasorb[®] (Curasan AG), a beta tricalcium phosphate ceramic, as a carrier substance for antibiotics by measuring *in vitro* the release kinetics of gentamicin and vancomycin [17, 18].

Former studies showed the principal usage of beta-tricalcium phosphate as a carrier substance for proteins like BMP-2 [19].

2. MATERIALS AND METHODS

2.1. Antibiotic Carrier

We used Cerasorb[®] and Cerasorb M[®] as it was supplied to us by the manufacturer Curasan AG, Germany. Cerasorb[®] ortho has an open interconnecting microporosity of 35 % and is available in a round shape. Cerasorb M[®] features an interconnecting, open multi porosity with micro, meso and macro pores (5 µm - 500 µm) and a total porosity of approx. 65%. The granules are polygonal, i.e. irregularly shaped, and facilitate canting and interlocking in the defect cavity. Micro movements are largely prevented.

Both materials have a phase purity of more than 99% β-tricalcium phosphate. For this study we had 10 pieces of Cerasorb[®] and Cerasorb M[®] at our disposal. Each of which measured 1x1x1 cm. One granule of each group was tested on the absorption of the antibiotics vancomycin and gentamicin. The remaining granules were tested on the

release kinetic either of vancomycin or gentamicin, 4 granules per each antibiotic.

2.2. Antibiotic Uptake and Release

Both bone graft substitutes were loaded with antibiotics *via* a soaking process. 5 pellets of Cerasorb[®] and Cerasorb M[®] were soaked for 1 minute in a gentamicin (Gentamicin-ratiopharm 80 SF, Ratiopharm GmbH, Ulm) solution (40 mg/ml). The pellets were kept at room temperature and completely covered by the solution during the soaking process.

The vancomycin solution was produced by injecting 10 ml water for injection into the dry matter vancomycin (Vancomycin CP, Hikma Pharma GmbH, Gräfeling), thus creating a solution of 50 mg/ml. 5 pellets of Cerasorb[®] and Cerasorb M[®] were soaked for 1 minute in the vancomycin solution. As above the pellets were completely covered by the solution and kept at room temperature during the soaking process.

To measure the antibiotic uptake one pellet of Cerasorb[®] and Cerasorb M[®] loaded with either one of the antibiotics was pestled. Subsequently the material was mixed with 4 ml PBS and incubated at 37° for 24 hours. Supernatants were removed and stored at -80°. The antibiotic content was measured in the supernatant.

In order to determine the release kinetics each of the two antibiotics was soaked into 4 pellets of Cerasorb[®] and 4 pellets of Cerasorb M[®]. We examined the elution in 4 ml phosphate-buffered saline (PBS) at pH 7,4 and 37°. Every 24 hours the supernatant was taken from the solution and 1 ml was preserved at -80°. The remaining pellets were cleaned with PBS and again covered with 4 ml of fresh PBS. This procedure was continued for 10 days.

The taken samples were preserved at -80° and defrosted immediately before being assayed with either MULTIGENT Gentamicin or Vancomycin assay (Abbott Laboratories Inc., USA), both homogeneous particle-enhanced turbidimetric inhibition immunoassays (PETINIA). Both assays were based on competition between drug in the sample and drug coated onto a microparticle for antibody binding sites of the gentamicin or vancomycin antibody reagent. The gentamicin- or vancomycin-coated microparticle reagent was rapidly agglutinated in the presence of the anti-gentamicin/vancomycin antibody reagent and in the absence of any competing drug in the sample.

The rate of absorbance change was measured photometrically and was directly proportional to the rate of agglutination of the particles. When a sample containing gentamicin/vancomycin was added, the agglutination reaction was partially inhibited, slowing down the rate of absorbance change. A concentration-dependent classic agglutination inhibition curve can be obtained with maximum rate of agglutination at the lowest gentamicin/vancomycin concentration, and the lowest agglutination rate at the highest gentamicin/vancomycin concentration.

The antibiotic concentration of the eluates was determined by an automatic antibiotic concentration analysing system (Architect 4000, Abbott Laboratories Inc., USA).

3. RESULTS

The loading of the granules showed no technical difficulties and does not affect the macroscopic or mechanical features of the granules after being loaded.

Antibiotic uptake of gentamicin was 406,38 µg/ml for Cerasorb[®] and 145,2 µg/ml for Cerasorb M[®]. The uptake of vancomycin was 1552,4 µg/ml for Cerasorb[®], Cerasorb M[®] granules had an uptake of 149 µg/ml.

Release kinetics of both granule types were measured as described by Rauschmann *et al.* [14]. Initially both materials showed a high release of the loaded antibiotics, Cerasorb M[®] showing a lower initial release level for gentamicin and vancomycin than Cerasorb[®]. At day 4 the gentamicin concentration of the Cerasorb M[®] granules was under the detection threshold, for Cerasorb[®] the gentamicin concentration was undetectable at day 6 (Fig. 1).

The vancomycin release-level followed a similar pattern, although the vancomycin concentration eluted by Cerasorb M[®] granules stayed above the detection threshold during the period of the experimental (Fig. 2).

DISCUSSION

Osteomyelitis is a difficult infection to treat and to eradicate. It represents a challenge for orthopaedic surgeons worldwide. Multiple surgical debridements and long-term parental antibiotics are often required for effective therapy. Besides these techniques, the local administration of antibiotics plays a crucial role in a successful therapy.

The main goal for local antibiotic therapies is to deliver an effective antimicrobial at sufficiently high concentrations to the area of infection. It is usually performed in addition with the use of polymethylmetacrylate (PMMA) beads and bone cements. The disadvantages associated with the usage of PMMA beads include being determined to one antibiotic at a time and the necessity of a second surgery to remove the beads after the antibiotic elution [2, 3, 21].

All these disadvantages made an intense search for possible alternatives necessary. An ideal antibiotic delivery system should provide an adequate antimicrobial concentration at the target site a constant release of antimicrobial over a prolonged period and be biodegradable to avoid the necessity of a second surgery [30].

One possible alternative is calciumsulphate, which is, as several studies showed, an adequate local drug delivery system [11, 12]. Admittedly during the first 60 days after implantation calciumsulphate causes inflammatory reactions in the circumjacent tissue [8]. After day 60 the inflammation ceases in the affected bone, but not in the circumjacent soft tissue.

Lactic-acid polymer gained interest as antibiotic carrier material. These polymers are biodegradable, making a second surgery to remove the carrier material unnecessary. *In vitro*, as well as *in vivo*, lactic-acid polymers showed a high release kinetic of loaded antibiotics [22-25]. A big drawback is its nonexistent osteoconductive property. Lactic-acid polymers are not commercially available. The addition of nanocrystalline hydroxyapatite to

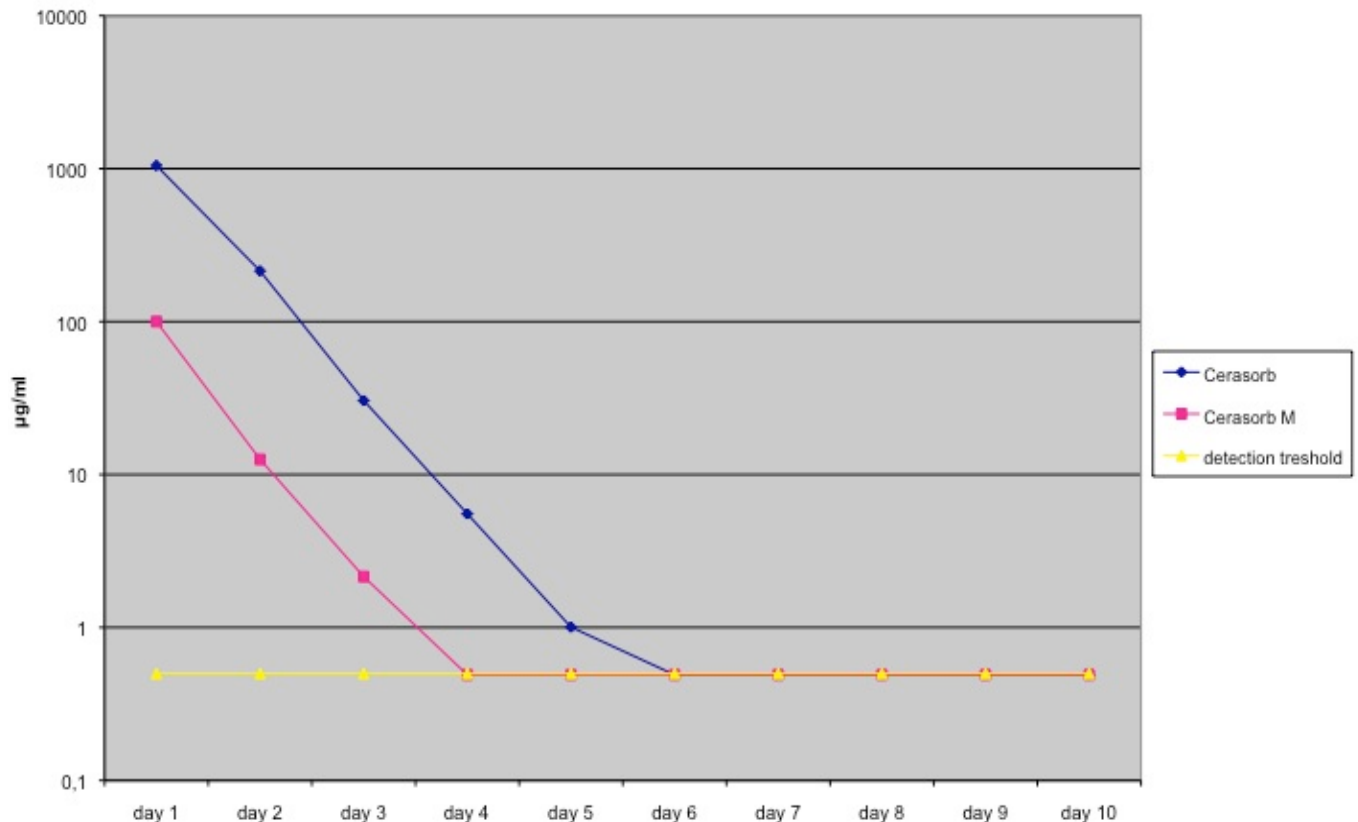


Fig. (1). Elution pharmacokinetics of Gentamicin from Cerasorb[®] and Cerasorb M[®]. Antibiotic release is expressed in µg/ml. Mean values were derived from a total of 4 different tests. Granules were loaded in a solution of 40 g/L antibiotics.

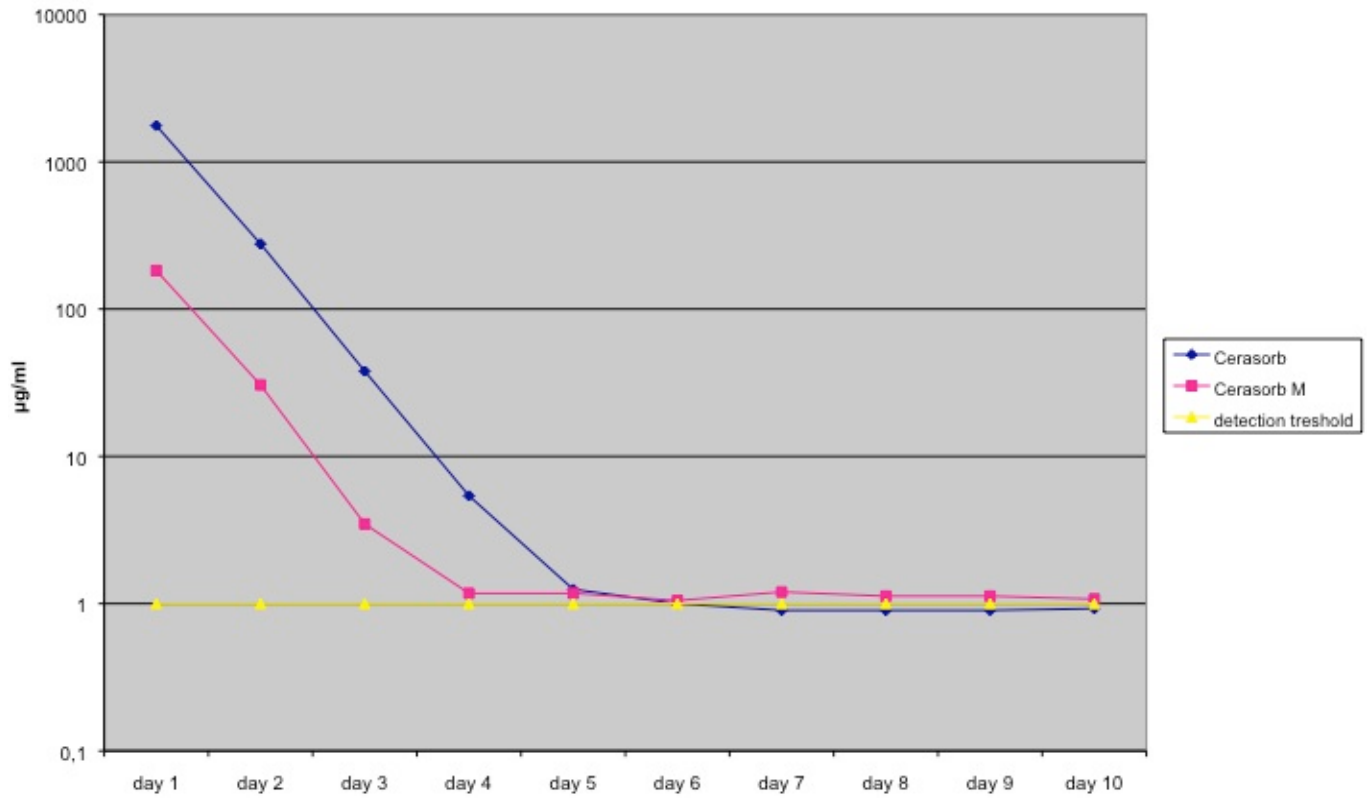


Fig. (2). Elution pharmacokinetics of vancomycin from Cerasorb[®] and Cerasorb M[®]. Antibiotic release is expressed in µg/ml. Mean values were derived from a total of 4 different tests. Granules were loaded in a solution of 50 g/L antibiotic.

calciumsulphate enhanced the biocompatibility of calciumsulphate in a composite material. This combination showed a reduced inflammatory reaction in the adjacent soft tissue in comparison to pure calciumsulphate, as well as higher osteoconductive properties and provides a scaffold for new bone formation [26]. *In vitro* the calciumsulphate hydroxyapatite composite showed a high release rate of the loaded antibiotics, in particular during the first days of the trial this antibiotic delivery system showed local effective antibiotic concentrations above the minimal inhibiting concentration (MIC) of vancomycin and gentamicin susceptible pathogens [14]. The most relevant pathogen for bone infections is Staph. Aureus, showing an MIC90 (antibiotic concentration that inhibits growth of 90% Staph. Aureus strains) of 1 mg/L for both vancomycin and gentamicin in susceptible strains [14, 23, 27].

Calcium sulphate preloaded with Tobramycin (OsteoSet^{®T}) in combination with surgical debridement showed promising results in the treatment of infected bone defects. However, an individualized chemotherapy in terms of local administration of different antibiotics is not provided [12]. To avoid the emerging tobramycin resistance custom-made calciumsulphate pellets, individually loaded with the appropriate antibiotic, are commercially available. In clinical and experimental studies these pellets showed a high efficiency in treating osteomyelitis. However, antibiotic loading has to be done before hardening and sterilization, creating a potential risk of reducing antibiotic efficiency. Adding the antibiotic during the process of hardening may impair the antibiotic elution. Dacquet *et al.* showed that the elution of teicoplanin, which was added to the calcium

sulphate powder before hardening, was only 1-5 µg during the first 10 days, too low for an effective osteomyelitis treatment [11]. Furthermore, there is a potential deceleration in the surgical procedure due to the 12-14 minutes of hardening.

In the current study we tried to minimize possible risks of impairing antibiotic activity due to any hardening or sterilization processes. To prevent inactivation of antibiotics by hardening or sterilization we performed the loading of antibiotics after these procedures. The ready-for use pellets completely absorbed the antibiotic solution. By comparing Cerasorb[®] and Cerasorb M[®] with their different porosities, we proved that antibiotic uptake and release are influenced by different porosities of the bone graft pellets. The higher porosity of Cerasorb M[®] showed a lower release of both loaded antibiotics from the start. The antibiotic concentrations never reached the antibiotic concentrations eluted by Cerasorb[®]. This result is in accordance with several other studies [14, 20]. Based on a minimal inhibition concentration (MIC) of 1 mg/L for both vancomycin and gentamicin, both materials achieved effective local antibiotic concentrations for Staph aureus within the first 4 and 5 days, respectively. After day 4 and 5, respectively, for both tested bone graft materials local antibiotic concentrations were less than the MIC and under the detection threshold. Rauschmann *et al.* reported similar results for the elution of vancomycin and gentamicin with nanocrystalline hydroxyapatite calcium sulphate as a carrier material. Local antibiotic concentrations for both antibiotics were initially high and stayed above the detection threshold for 3-4 days [14]. The present study showed concentrations of released gentamicin and

Table 1. Elution Pharmacokinetics and Standard Deviation of Gentamicin from Cerasorb® and Cerasorb M®. Antibiotic Release is Expressed in µg/ml. Mean Values were Derived from a Total of 4 Different Tests. Granules were Loaded in a Solution of 40 g/L Antibiotics

	Elution Kinetics of Gentamicin									
	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7	Day 8	Day 9	Day 10
Cerasorb	1046.4675	214.3825	30.475	5.5325	1.005	0.49	0.49	0.49	0.49	0.49
Standard Deviation	421.3	18.8	3.4	0.61	0.26	0	0	0	0	0
Cerasorb M	99.9675	12.555	2.15	0.49	0.49	0.49	0.49	0.49	0.49	0.49
Standard Deviation	34.8	3.2	0.9	0	0	0	0	0	0	0

Table 2. Elution Pharmacokinetics and Standard Deviation of Vancomycin from Cerasorb® and Cerasorb M®. Antibiotic Release is Expressed in µg/ml. Mean Values were Derived from a Total of 4 Different Tests. Granules were Loaded in a Solution of 50 g/L Antibiotics

	Elution Kinetics of Vancomycin									
	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7	Day 8	Day 9	Day 10
Cerasorb	1761.3	277.35	37.875	5.4	1.25	1	0.9	0.9	0.9	0.925
standard deviation	331.87	32.46	2.55	0.41	0.25	0.2	0	0	0	0.05
Cerasorb M	182.775	30.55	3.475	1.175	1.175	1.05	1.2	1.125	1.125	1.075
standard deviation	27.9	23.4	0.56	0.4	0.3	0.3	0.38	0.45	0.45	0.35
detection treshold	1	1	1	1	1	1	1	1	1	1

vancomycin exceeding 100-1000-fold the MIC of susceptible *Staph. aureus* within the first day, at day 2 and 3 the MIC was exceeded 10-100-fold.

Prima facie these results seem to be comparable with the results reported by Rauschmann *et al.*, however, the main difference is the method to measure the antibiotic concentration. Rauschmann *et al.* used a standard agar diffusion test with *Bacillus subtilis* ATCC 6633. In contrast, we used machine-aided diagnosis to evaluate the antibiotic elution of both used bone graft substitutes [14]. Wichelhaus *et al.* used the standard agar diffusion test as well in their evaluation of the release of 4 different antibiotics from calcium sulphate carrier beads and found antibiotic release to be above the MIC for a period of ten days [20].

Miclau *et al.* showed a high initial antibiotic release from autogenic bone graft and demineralized bone matrix during the first days of their study, whereas PMMA- and calcium sulphate carrier beads showed a prolonged release of the loaded antibiotics [29]. A major drawback of autogenic bone graft is the required removal of the bone graft resulting in a high risk for complications, as well as its limited disposability. Demineralized bone matrix is not rugged and due to its structure is limited in the way it acts as an osteoconductive scaffold.

In contrast calcium phosphate ceramics, like hydroxyapatite and tricalciumphosphate, are available in large quantities and show excellent osteoconductive features. Different studies have proven the qualification of hydroxyapatite as a local antibiotic delivery system [28, 30].

CONCLUSION

The tricalcium phosphate ceramics Cerasorb® and Cerasorb® M are well established as bone graft substitutes. In the current study we loaded both ceramics with vancomycin and gentamicin to create a local antibiotic delivery system. The pre-fabricated pellets were loaded after sterilization and hardening to prevent inactivation of antibiotics by these procedures. We showed a high initial antibiotic release with subsequent decline well above the MIC of *Staph aureus* for both materials within the first 4 and 5 days, respectively. The pre-fabricated pellets offer fast and technically easy loading with the selected antibiotic. Loading can be done according to antibiograms offering individualized antibiotic treatment options. Due to the osteoconductive and biodegradability properties of Cerasorb® and Cerasorb M® there is no need for removal of the material as in the case with PMMA. Given the different technical features of both ceramics it enables practitioners to react to different clinical requirements, e.g. slower or faster resorption.

In conclusion Cerasorb® and Cerasorb M® has potential as a new treatment option in osteomyelitis and infectious bone disease.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

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REFERENCES

- [1] Harle A, Ritzerfeld J. The release of gentamycin into the wound secretions from polymethylmethacrylate beads. A study with reference to the animal experiment. *Arch Orthop Trauma Surg* 1979; 95 (1-2): 65-70.
- [2] Klemm K. Gentamicin-PMMA-beads in treating bone and soft tissue infections (author's transl). *Zentralbl Chir* 1979; 104(14): 934-42.
- [3] Buchholz HW, Engelbrecht H. [Depot effects of various antibiotics mixed with Palacos resins]. *Chirurg* 1970; 41(11): 511-5.
- [4] Alexander DL, Manson NA, Mitchell MJ. Efficacy of calcium sulfate plus decompression bone in lumbar and lumbosacral spinal fusion: preliminary results in 40 patients. *Can J Surg* 2001; 44(4): 262-6.
- [5] Gitelis S, Piasecki P, Turner T, Haggard W, Charters J, Urban R. Use of a calcium sulfate-based bone graft substitute for benign bone lesions. *Orthopedics* 2001; 24(2): 162-6.
- [6] Kelly CM, Wilkins RM, Gitelis S, Hartjen C, Watson JT, Kim PT. The use of a surgical grade calcium sulfate as a bone graft substitute: results of a multicenter trial. *Clin Orthop Relat Res* 2001; (382): 42-50.
- [7] Mirzayan R, Panossian V, Avedian R, Forrester DM, Menendez LR. The use of calcium sulfate in the treatment of benign bone lesions. A preliminary report. *J Bone Joint Surg Am* 2001; 83-A(3): 355-8.
- [8] Coetzee AS. Regeneration of bone in the presence of calcium sulfate. *Arch Otolaryngol* 1980; 106(7): 405-9.
- [9] Lee GH, Khoury JG, Bell JE, Buckwalter JA. Adverse reactions to OsteoSet bone graft substitute, the incidence in a consecutive series. *Iowa Orthop J* 2002; 22: 35-8.
- [10] Robinson D, Alk D, Sandbank J, Farber R, Halperin N. Inflammatory reactions associated with a calcium sulfate bone substitute. *Ann Transplant* 1999; 4(3-4): 91-7.
- [11] Dacquet V, Varlet A, Tandogan RN, *et al.* Antibiotic-impregnated plaster of Paris beads. Trials with teicoplanin. *Clin Orthop Relat Res* 1992; 282: 241-9.
- [12] McKee MD, Wild LM, Schmeimtsch EH, Waddell JP. The use of an antibiotic-impregnated, osteoconductive, bioabsorbable bone substitute in the treatment of infected long bone defects: early results of a prospective trial. *J Orthop Trauma* 2002; 16(9): 622-7.
- [13] Gitelis, S, Brebach, GT. The treatment of chronic osteomyelitis with a biodegradable antibiotic-impregnated implant. *J Orthop Surg (Hong Kong)* 2002; 10(1): 53-60.
- [14] Rauschmann MA, Wichelhaus TA, Stirnal V, *et al.* Nanocrystalline hydroxyapatite and calcium sulphate as biodegradable composite carrier material for local delivery of antibiotics in bone infections. *Biomaterials* 2005; 26: 2677-84.
- [15] Dorozhkin SV, Epple M. Biological and medical significance of calcium phosphates. *Angew Chem Int Ed Engl* 2002; 41(17): 3130-46.
- [16] Ransford AO, Morley T, Edgar MA, *et al.* Synthetic porous ceramic compared with autograft in scoliosis surgery. A prospective, randomized study of 341 patients. *J Bone Joint Surg Br* 1998; 80(1): 13-8.
- [17] Szabo G, Huys L, Coullthard P, *et al.* A prospective multicenter randomized clinical trial of autogenous bone versus beta-tricalcium phosphate graft alone for bilateral sinus elevation: histologic and histomorphometric evaluation. *Int J Oral Maxillofac Implants* 2005; 20(3): 371-81.
- [18] Maus U. Klinische Erfahrungen mit dem Knochenersatzstoff Cerasorb. *Orthopädische Praxis* 2007; 43: 258-61.
- [19] Kim CS, Kim JI, Kim J, Choi SH, Chai JK, Kim CK, Cho KS. Ectopic bone formation associated with recombinant human bone morphogenetic proteins-2 using absorbable collagen sponge and beta tricalcium phosphate as carriers. *Biomaterials* 2005; 26(15): 2501-7.
- [20] Wichelhaus TA, Dingeldein E, Rauschmann M, *et al.* Elution characteristics of vancomycin, teicoplanin, gentamicin and clindamycin from calcium sulphate beads. *J Antimicrob Chemother* 2001; 48(1): 117-9.
- [21] Liu SJ, Ueng SW, Chan EC, *et al.* *In vitro* elution of vancomycin from biodegradable beads. *J Biomed Mater Res* 1999; 48(5): 613-20.
- [22] Benoit MA, Mousset B, Delloye C, Bouillet R, Gillard J. Antibiotic-loaded plaster of Paris implants coated with poly lactide-co-glycolide as a controlled release delivery system for the treatment of bone infections. *Int Orthop* 1997; 21(6): 403-8.
- [23] Kanellakopoulou K, Kolia M, Anastasiadis A, *et al.* Lactic acid polymers as biodegradable carriers of fluoroquinolones: an *in vitro* study. *Antimicrob Agents Chemother* 1999; 43(3): 714-6.
- [24] Kanellakopoulou K, Galanakis N, Giamarellos-Bourboulis EJ, *et al.* Treatment of experimental osteomyelitis caused by methicillin-resistant *Staphylococcus aureus* with a biodegradable system of lactic acid polymer releasing pefloxacin. *J Antimicrob Chemother* 2000; 46(2): 311-4.
- [25] Wie G, Kotoura Y, Oka M, *et al.* A bioabsorbable delivery system for antibiotic treatment of osteomyelitis. The use of lactic acid oligomer as a carrier. *J Bone Joint Surg Br* 1991; 73(2): 246-52.
- [26] Sato S, Koshino T, Saito T. Osteogenic response of rabbit tibia to hydroxyapatite particle-Plaster of Paris mixture. *Biomaterials* 1998; 19(20): 1895-900.
- [27] Fluit AC, Jones ME, Schmitz FJ, Acar J, Gupta R, Verhoef J. Antimicrobial susceptibility and frequency of occurrence of clinical blood isolates in Europe from the SENTRY antimicrobial surveillance program, 1997 and 1998. *Clin Infect Dis* 2000; 30(3): 454-60.
- [28] Cornell CN, Tyndall D, Walter S, Lane JM, Brause BD. Treatment of experimental osteomyelitis with antibiotic-impregnated bone graft substitute. *J Orthop Res* 1993; 11(5): 619-26.
- [29] Miclau T, Dahners LE, Lindsey RW. *In vitro* pharmacokinetics of antibiotic release from locally implantable materials. *J Orthop Res* 1993; 11(5): 627-32.
- [30] Joosten U, Joist A, Gosheger G, Liljenqvist U, Brandt B, von Eiff C. Effectiveness of hydroxyapatite-vancomycin bone cement in the treatment of *Staphylococcus aureus* induced chronic osteomyelitis. *Biomaterials* 2005; 26(25): 5251-8.

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