

# Application of Autologous Bone Marrow Derived Mesenchymal Stem Cells to an Ovine Model of Growth Plate Cartilage Injury

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**Abstract:** Injury to growth plate cartilage in children can lead to bone bridge formation and result in bone growth deformities, a significant clinical problem currently lacking biological treatment. Mesenchymal stem/stromal cells (MSC) offer a promising therapeutic option for regeneration of damaged cartilage, due to their self renewing and multi-lineage differentiation attributes. Although some small animal model studies highlight the therapeutic potential of MSC for growth plate repair, translational research in large animal models, which more closely resemble the human condition, are lacking. Our laboratory has recently characterised MSCs derived from ovine bone marrow, and demonstrated these cells form cartilage-like tissue when transplanted within the gelatin sponge, Gelfoam, *in vivo*. In the current study, autologous bone marrow MSC were seeded into Gelfoam scaffold containing TGF- $\beta$ 1, and transplanted into a surgically created defect of the proximal ovine tibial growth plate. Examination of implants at 5 week post-operatively revealed transplanted autologous MSC failed to form new cartilage structure at the defect site, but contributed to an increase in formation of a dense fibrous tissue. Importantly, the extent of osteogenesis was diminished, and bone bridge formation was not accelerated due to transplantation of MSCs or the gelatin scaffold. The current study represents the first work that has utilised this ovine large animal model to investigate whether autologous bone marrow derived MSC can be used to initiate regeneration at the injured growth plate.

**Keywords:** Mesenchymal stem cell, bone marrow, ovine, growth plate, tissue engineering.

## INTRODUCTION

The identification of self renewing multipotential progenitor cells from adult stromal tissues has stimulated significant interest and promise for the utilisation of these cells in regenerative medicine and tissue engineering. Since the discovery of putative mesenchymal stem/stromal cells (MSC) from bone marrow suspensions [1] with the ability to form adherent clonogenic clusters (CFU-F), and capacity to differentiate into a multitude of cell types *in vitro* and *in vivo*, MSCs have now been isolated from numerous connective tissues [2]. In addition, MSCs appear to share similar characteristics across species, which has facilitated the application of MSC in translational studies using animal models.

The growth plate is a unique cartilaginous tissue located at the proximal and distal ends of the long bones of children, and is responsible for longitudinal bone growth until closure at skeletal maturity. Due to the avascular and alymphatic nature of cartilage, once damaged, cartilage has a poor

regenerative capacity. Injury to growth plate cartilage often results in an undesirable repair response mechanism at the site of injury, where ossification of the damaged tissue may lead to formation a bone bridge. Establishment of a bone bridge across the growth plate can have serious consequences in children, and may lead to limb length discrepancies and angular deformity. Current treatments of limb abnormalities include surgical correction of angularity or length discrepancy after manifestation of the deformity. To date, there is no cell-based biological based therapy to prevent bone bridge formation and regenerate the damaged growth plate cartilage in clinical practice [2].

Although there have been some recent investigations in rabbit or rodent models examining potential applications of MSCs in repairing injured growth plate with some success [3-6], the therapeutic potential of MSCs in growth plate regeneration remains to be investigated in a large animal model. The use of sheep as a large animal model for orthopaedic research continues to increase in popularity due to similarities with humans in weight, size, joint structure, bone/cartilage regenerative processes, and thus the potential in translational research.

Our laboratory has undertaken numerous studies employing a sheep model to investigate mechanisms of

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growth plate injury repair, techniques for the prevention of bone bridge formation, and reversal of skeletal deformity [7-13]. Several interpositional materials have been assessed for their potential to impede or prevent bone formation and regenerate the damaged cartilage. These include fat, cultured chondrocytes [7, 9], cartilage [11], periosteum [11], and a type 1 collagen paste [12].

We have recently described the functional and phenotypic properties of ovine bone marrow derived MSCs, including incidence, immunophenotype, proliferative response to mitogens, and differentiation potential both *in vitro* and *in vivo* [14]. In this pilot study the potential of autologous MSCs to repair damaged growth plate cartilage in an immature ovine model was investigated.

## MATERIALS AND METHODS

### Subjects and Cell Culture

All procedures were approved by the institutional animal ethics committee and performed under sterile conditions. Fresh bone marrow aspirates were obtained from the iliac crest of 5 lambs (approximately 8 weeks old) under general anaesthesia according to procedures approved by the ethics committee of the Women's & Children's Hospital, South Australia. Bone marrow mononuclear cells (BM MNC) were isolated by centrifugation of aspirates on a Lymphoprep™ density gradient (S.G. 1.077 g/ml) (Axis Shield or Nycomed, Oslo, Norway) as previously described [15, 16].

### Ovine Primary Mesenchymal Stem Cell Cultures

Briefly, single cell suspensions of BM MNC were plated in monolayer in  $\alpha$ -MEM media supplemented with penicillin (50 i.u./ml)/streptomycin sulphate (50  $\mu$ g/ml), 10% (v/v) fetal calf serum (FCS; SAFC Biosciences, Lenexa, KS), 2 mM L-glutamine (SAFC Biosciences), 1 mM sodium pyruvate (SAFC Biosciences), and 100  $\mu$ M L-ascorbate-2-phosphate (Asc-2-P, WAKO Pure Chemical Industries Ltd, Osaka, Japan) and incubated at 37°C in the presence of 5% CO<sub>2</sub>. After 3 days, non adherent cells were removed and fresh media replaced. Upon confluence, cells were detached by enzymatic digestion in 0.05% (w/v) trypsin (Gibco, Invitrogen, Carlsbad, CA, USA) and 0.5 mM EDTA (BDH AnalAR®, Merck) in sterile PBS.

### Transplantation of Autologous MSC to Growth Plate Defect in Lambs

Four million lamb MSCs (P1) were resuspended in serum replete chondrogenic media containing growth factor TGF- $\beta$ 1 (10 ng/ml) and statically seeded onto Gelfoam sponge (Pharmacia & Upjohn) (approximately 10 x 8 mm squares). The MSC were consolidated within the Gelfoam sponge using a fibrin clot (gift from Mr. Rick Tocchetti, IMVS, Adelaide, Australia).

The procedure for creation of a small (10 mm deep x 10 mm wide x 5 mm high) growth plate defect at the proximal tibia of lambs has been established previously by Foster *et al.* [9, 13]. The defect site was immersed in 100  $\mu$ l of 1 Unit/ml Chondroitinase ABC (Sigma-Aldrich), which was removed by aspiration and blotted dry after 5 mins. The Gelfoam scaffold with or without MSCs was placed within the defect site. Nine lambs were divided into two groups. One group (n=5) received a growth plate defect on both hind

legs, with one defect on one limb being filled with Gelfoam scaffold in chondrogenic media containing no cells, with the defect on the contralateral limb receiving the Gelfoam scaffold seeded with 4 x 10<sup>6</sup> autologous MSC. The other group (n = 4) received an untreated defect on one limb and the contralateral limb was uninjured as a normal control. Two titanium Kirschener wires (K-wires) were inserted after wound closure perpendicular to the growth plate, in the epiphysis and diaphysis respectively, using a F-shaped template of 20 mm separation, in order to facilitate the precise measurement of limb growth following partial growth plate disturbance.

All animals were sacrificed five weeks post-operatively. In order to perfuse, fix, and collect tissue specimens, animals were sedated and general anaesthesia was induced as previously described [9, 13]. Following catheterisation the leg was perfused with 500ml of 1% sodium nitrite in 0.1 M phosphate buffer. During the sodium nitrite perfusion, the animal was euthanized by an overdose of sodium pentobarbitone (10ml, Lethabarb, Virbac, Milperra, NSW, Australia). Subsequently, each leg was perfused with 500 ml of 10% buffered formalin. The hind leg was removed and the portion (15 mm x 15 mm x 15 mm) containing the defect was excised using an Isomet Low Speed Saw (Buehler Ltd., Lake Bluff, IL, USA) and placed in a solution of 10% buffered formalin for 48 hours. Following fixation, the sample was decalcified in Immunocal (United Biosciences, Carindale, Qld, Australia) for 7 days and processed for paraffin embedding, from which 5  $\mu$ m sections were cut and stained with Haematoxylin, Eosin, and 1% Alcian Blue.

### Growth Plate Injury Repair Measurements and Statistical Analysis

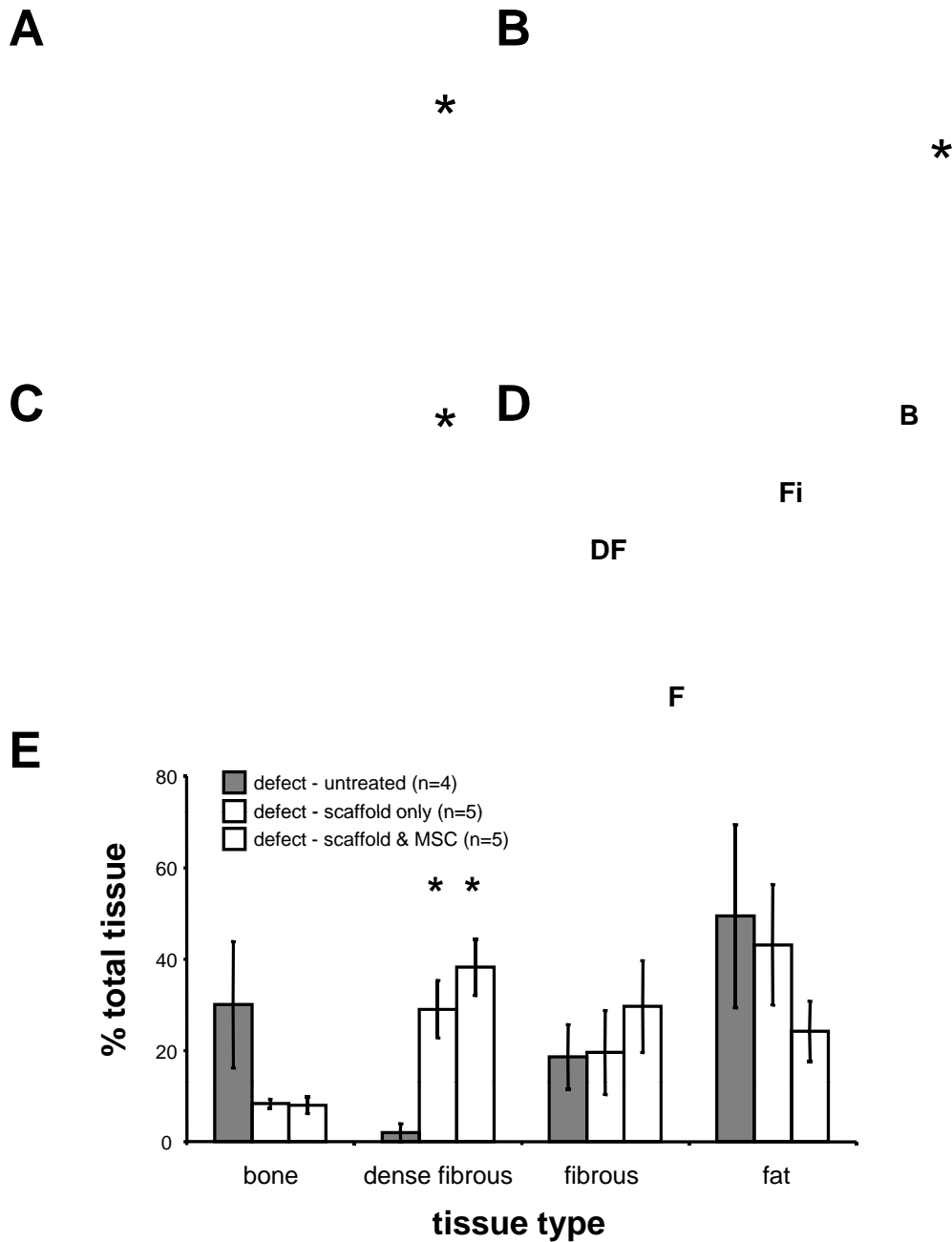
Measurement of the area representative of different repair tissue types (mesenchymal, bone, cartilage, marrow, and fat) within lamb growth plate defects was performed using the Olysia BioReport software imaging system (Olympus Corporation, Tokyo, Japan). Measurements were taken within a defined area adjacent to the intact growth plate and calculated as the percentage of the total area measured [17]. The measurements for each group were combined and compared for statistical significance using either a paired t-test or for non parametric data, the Mann-Whitney test. Statistical significance was confirmed where  $p < 0.05$ .

## RESULTS

Morphologically, ovine MSC are a large adherent population exhibiting a heterogeneous phenotype including cells resembling fibroblasts, polygonal, stellate, and spindle shaped cells with long processes (Fig. 1A). Gelfoam, a medical device made from porcine skin gelatin and used clinically, was used as a scaffold carrier for MSC implantation in this study (Fig. 1B). Merino cross lambs (8 to 10 weeks old) were placed under general anaesthesia, and using a high speed dental drill and small dental burr (2 mm), a single partial peripheral growth plate defect was created of approximately 1 cm<sup>2</sup> (Fig. 1C). In the instance of transplantation with scaffold or scaffold plus autologous MSC, the Gelfoam implant was easily placed and fit into the defect site, and due to the absorbent and spongy characteristic of the scaffold, was highly compatible with the topography of the defect (Fig. 1D).







**Fig. (3).** Surgical creation of a growth plate defect and analysis of repair tissues 5 weeks after untreated or treated with interpositional transplantation of Gelfoam scaffold alone or seeded with autologous BM MSC. Representative images of repair tissues on cross sections stained with Haematoxylin, Eosin, and Alcian Blue (A-C). The location of the growth plate is indicated by \*. A representative example of extensive bone bridge formation is observed in the untreated defect (2x) (A). A representative example of growth plate defect treated with Gelfoam scaffold only (2x) (B), and defect treated with Gelfoam scaffold and autologous MSC (2x) (C). An example of different tissues observed within defect (4x) (D), which include bone (B), fat (F), fibrous tissue (Fi), and dense fibrous tissue (DF). Using Image Analysis software, the proportions of each repair tissue type were determined and are presented at the % mean area ± SEM of the total injury area for each group (E). Statistical significance between treatment groups is indicated by a \* (Mann-Whitney,  $p < 0.05$ ).

successful repair of growth plate defects using MSC has been achieved using agarose, chitin, gelatin and Gelfoam composites, and fibrin glue [3-6, 28]. The aforementioned studies all utilised a rabbit model, however the therapeutic potential of MSCs for growth plate regeneration in larger animal models has not been documented.

Recently, we demonstrated that adult ovine bone marrow aspirates contain a proportion of CFU-F cells with phenotypic and functional similarities to human MSC capable of multipotential differentiation *in vitro* and *in vivo* [14]. Undifferentiated ovine BM MSC seeded in Gelfoam with TGF-β1 form cartilage-like tissue, but not osteogenic

tissue, following subcutaneous transplantation in immunocompromised mice [14]. In the current study, Gelfoam scaffold was further selected for application in a model of growth plate injury for three additional reasons. Firstly, used clinically as a haemostatic agent, the intrinsic properties of Gelfoam may be beneficial in inhibiting the early inflammatory response that occurs following mechanical disruption of the growth plate, by physically impeding the infiltration of inflammatory cells. Secondly, the topography of Gelfoam renders it ideal for implantation in the defect created here using a lamb model. Thirdly, the characteristic of Gelfoam to withhold a high fluid content within its matrices may recapitulate cartilage tissue where the water content can extend to 80% of the wet weight. Gelfoam or gelatin sponges have been applied in studies examining chondrogenesis *in vivo* and cartilage repair, using both chondrocytes [29, 30] and MSC [3, 31-33].

The current study represents the first work that has utilised this ovine large animal model to investigate whether bone marrow derived MSC can be used to initiate regeneration at the injured growth plate. Results indicated that the application of MSC to the site of growth plate injury did not result in chondrogenic differentiation or regeneration of the damaged cartilage, but formation of dense fibrous tissue. The differentiation of cells to form more fibrous repair tissue may be attributed to the inflammatory response at the injury site, which may overwhelm signals imparted by the scaffold and the growth factor. Nonetheless, the undesirable bone formation was reduced and most importantly, not accelerated at the injury site. The application of scaffold without MSC reduced the proportion of bony tissue and the repair site contained fibrous tissue suggesting the infiltration of endogenous fibroblastic cells to the injury site. Taken together, the Gelfoam scaffold does not appear to be pro-osteogenic, which is desirable in studies addressing cartilage regeneration.

Questions to be addressed in the future may include 1) analysis of the repair tissue at longer intervals post-operatively to determine if the repair tissue is maintained and bone bridge is prevented long term, and thus whether different treatment groups are affecting reaction kinetics rather than overall outcome; 2) the potential of MSCs 'pre-differentiated/committed' to chondrogenic lineage *in vitro* prior to transplantation - to improve the possibility of chondrogenic tissue repair; 3) tracking of the introduced MSCs to determine their role in the repair response - whether MSCs directly contribute to the repair tissue or facilitate the recruitment of endogenous cells. Further study is required to evaluate whether MSC may be a viable therapeutic option for the biological regeneration of the growth plate following injury.

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